

# CHARACTERIZATION OF PROGRESSION OF GENOMIC CHANGES DURING CLINICAL COURSE OF AML



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BACKGROUND

## **NATURE OF ACUTE MYELOID LEUKEMIA**

role of bone marrow stem cells

insensitivity to chemotherapy



# ACUTE MYELOID LEUKEMIA

bone marrow malignancy

rapid growth of abnormal white cells which accumulate and interfere with production of normal blood cells

1,400 cases per year in Canada, 5-year survival 15-70%

## **INEFFECTIVE CLINICAL COURSE**

50% of patients relapse after chemotherapy and require bone marrow transplant

## **OPPORTUNITY FOR IMPROVING OUTCOMES**

little is known about reasons for relapse in normal karyotype AML

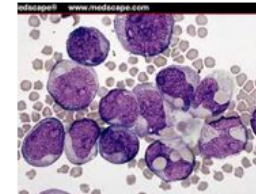
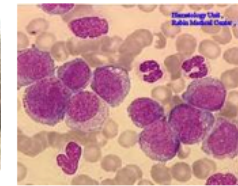
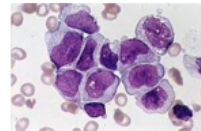
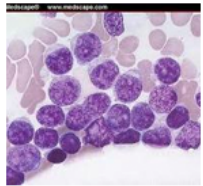
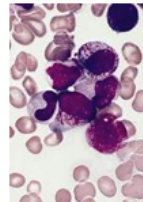
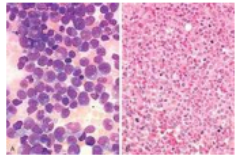
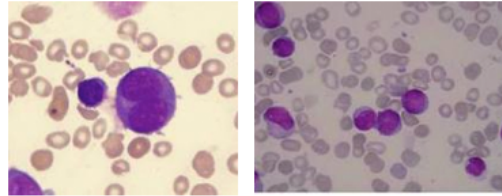
## **MODEL FOR GENOME EVOLUTION AND PROGRESSION**

bone marrow cells essentially eradicated during chemotherapy

population bottleneck amplifies effect of clonal expansion conferring resistance

# WHAT DOES AML LOOK LIKE?

AML	Incidence	Subtypes	Characteristics	Cytogenetics (karyotype)	Exacerbations
M1	2-5%	Acute Myeloid Leukemia (M1)	Myeloblasts	Normal	May be associated with t(8;21)
M2	20%	Acute Myeloid Leukemia (M2)	Myeloblasts	Normal	May be associated with t(8;21)
M3	3-5%	Acute Myeloid Leukemia (M3)	Myeloblasts	Normal	May be associated with t(11;17)
M4	10%	Acute Myeloid Leukemia (M4)	Myeloblasts	Normal	May be associated with t(8;21)
M5	2-5%	Acute Myeloid Leukemia (M5)	Myeloblasts	Normal	May be associated with t(8;21)
M6	1-2%	Acute Myeloid Leukemia (M6)	Myeloblasts	Normal	May be associated with t(8;21)
M7	1-2%	Acute Myeloid Leukemia (M7)	Myeloblasts	Normal	May be associated with t(8;21)
M8	1-2%	Acute Myeloid Leukemia (M8)	Myeloblasts	Normal	May be associated with t(8;21)



Google image searching for AML retrieves slides of cells ... and light-armoured vehicles (Panhard AML-90), already known to be ineffective in battle against AML

# CHEMOTHERAPY RESISTANCE - BONE MARROW STEM CELLS

bone marrow stem cells contribute to resistance to chemotherapy

quiescent

express transport proteins that expel toxins

## **RESISTANT SUBCLONES**

populations of cells with mutations that confer resistance

resistant subclones may be present at diagnosis or created by chemotherapy

differential mutational frequency may exist in stem cell compartment

RELEVANCE

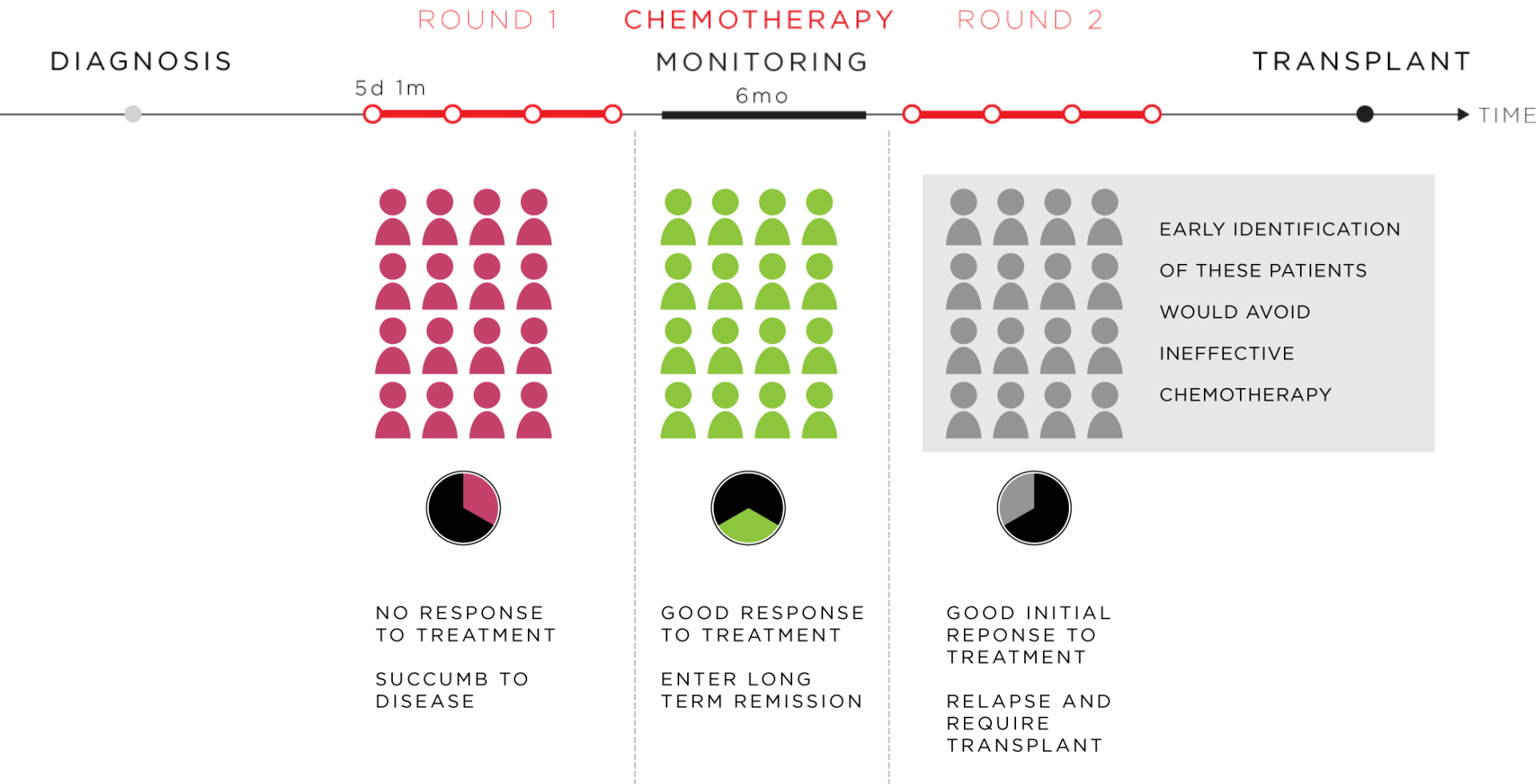
## **IMPROVED TREATMENT MANAGEMENT**

current prognosis methods lack specificity

standard treatment protocol is inefficient for many patients

# CURRENT TREATMENT METHODS

patients receive a combination of chemotherapeutics



## CURRENT PROGNOSIS METHODS

AML samples characterized with Sanger-type sequencing at diagnosis

detection limited to ~20% allele frequency

insensitive to rare subclones

no progression monitoring, to identify emergent resistant subclones



OBJECTIVE

## **IMPROVE AML TREATMENT OUTCOME**

characterize molecular evolution of disease during treatment

correlate genomic alterations with outcome

## **BETTER DISEASE TYPE CLASSIFIER**

identify prognostic markers and therapeutic targets



METHODS AND ANALYSIS

## **SAMPLE COLLECTION AND CHARACTERIZATION**

collect and separate bone marrow samples

sequence samples to identify new mutations

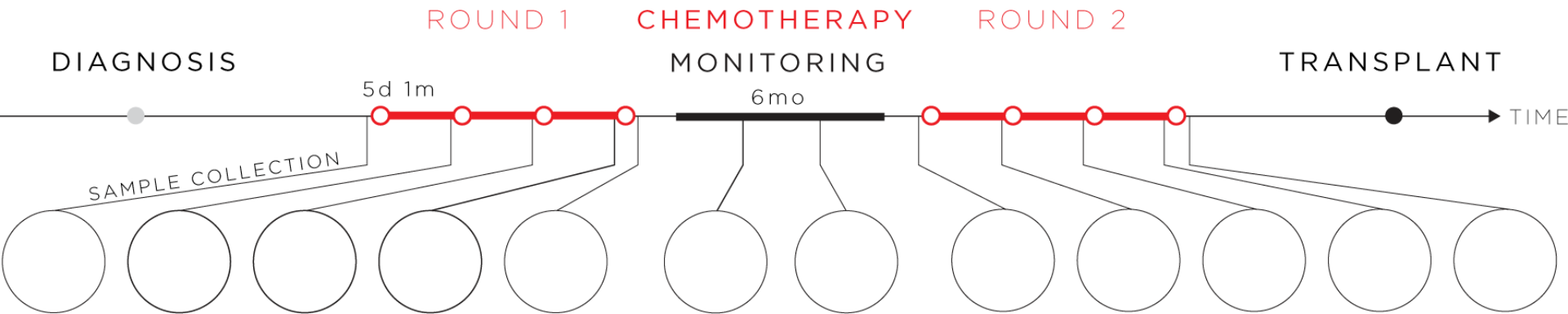
assemble genomes and transcriptomes

interrogate mutations in samples with low cell count using PCR

# SAMPLE COLLECTION

follow 30 normal karyotype AML patients from time of diagnosis

collect bone marrow samples prospectively during treatment and monitoring



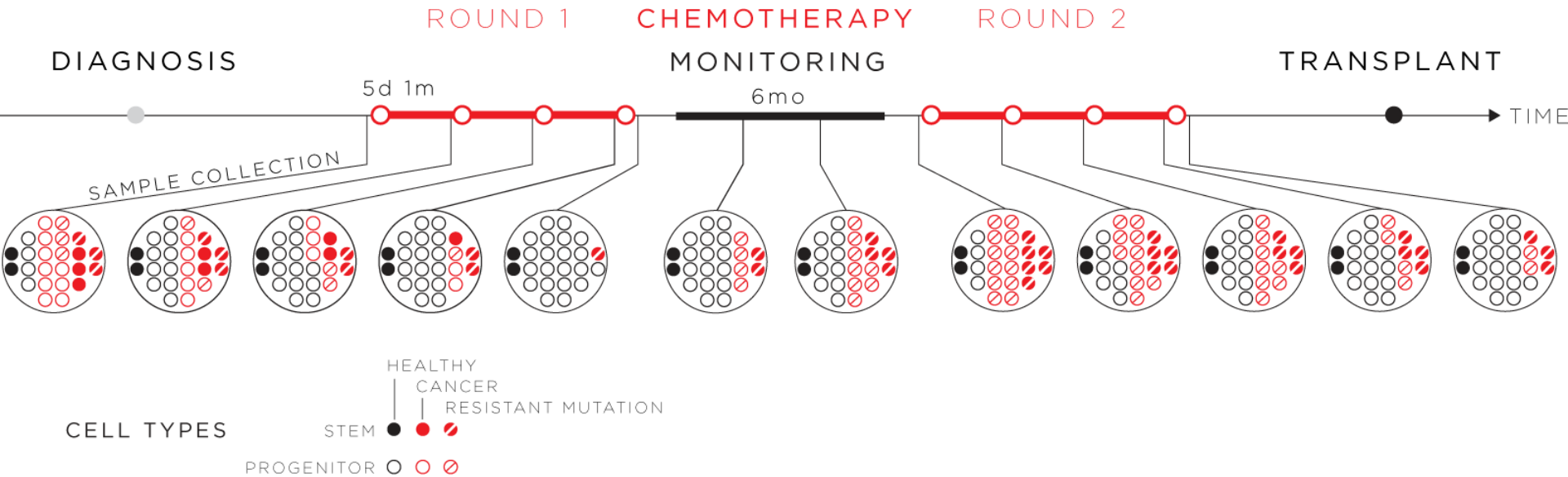
collect skin biopsy for matched normal

# WHAT DO WE EXPECT FROM RELAPSE CASES?

cancer cell population drastically reduced during chemotherapy

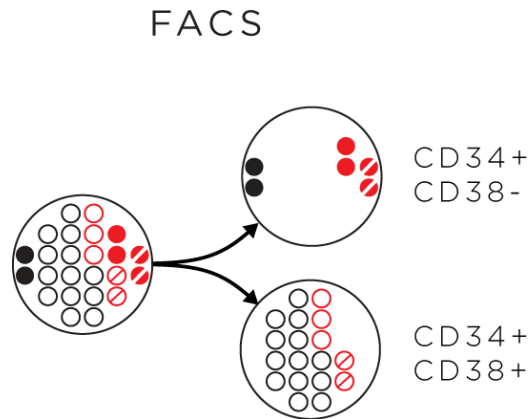
population increases during monitoring

rare resistant subclones dominate



## SEPARATE CELLS AND CREATE LIBRARIES

apply flow cytometry to separate stem cells from progenitors



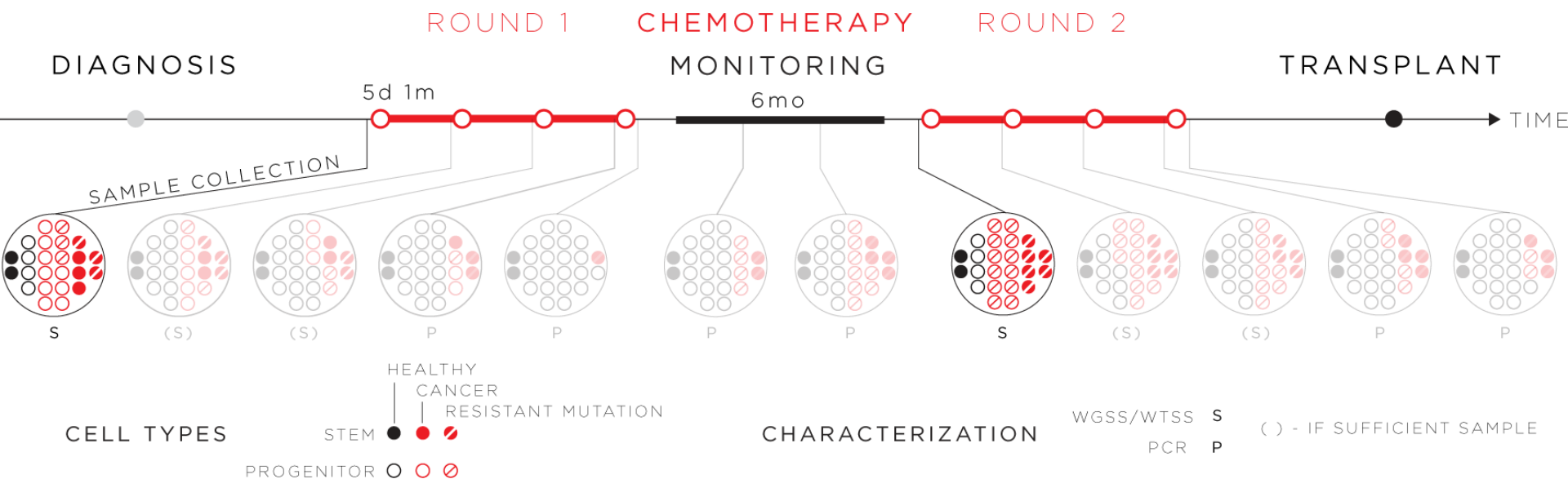
generate DNA and RNA libraries for WGSS and WTSS

### ROLE OF STEM CELL FRACTIONATE

genomic and gene expression landscape of stem cells will be different

# CHARACTERIZE SAMPLES

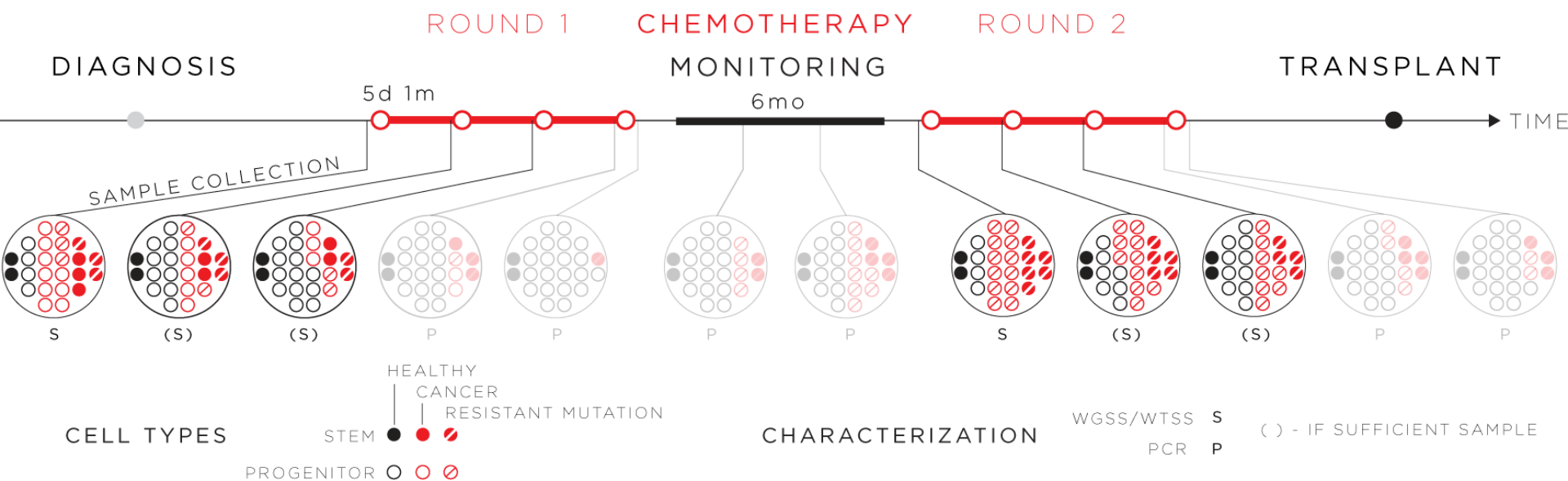
apply HiSeq to samples with sufficient cell count  
 at diagnosis  
 at relapse



# CHARACTERIZE SAMPLES

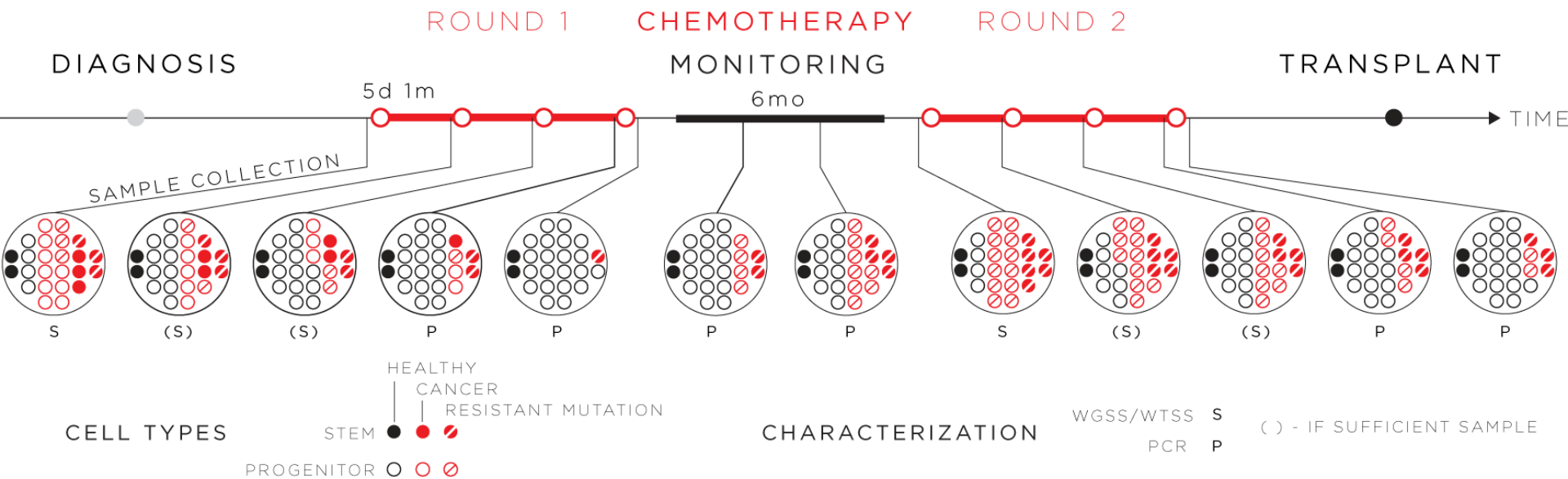
samples at 1<sup>st</sup>/2<sup>nd</sup> treatment in each round may have sufficient cell content

if so, sequence these



# CHARACTERIZE SAMPLES

interrogate remaining samples with PCR





# BIOINFORMATICS

## REFERENCE ALIGNMENT

align with BWA

identify SNV with samtools/SNVMix

use pair-end reads to map rearrangements

identify alternate splice sites with Tophat, HMMSplicer

perform expression and alternative expression analysis

## DE NOVO ASSEMBLY

assemble genome with ABySS

assemble transcriptome with Trans-ABYSS

identify fusion transcripts with deFuse

## CELL COUNTS AND ALLELE FREQUENCY

### LEUKEMIC STEM CELLS

expect  $0.1-10 \times 10^6$  LSC per sample at diagnosis and relapse  
this is sufficient DNA/RNA sequencing  
samples from which sufficient genomic material cannot be collected  
will be interrogated by PCR

### TARGETED DEEP SEQUENCING

during treatment, malignant cells will be regenerated by normal cells  
FACS may not reliably separate leukemic and normal cells due to novel phenotypes (new markers)  
to identify presence of rare alleles, we will apply deep targeted sequencing based on WGSS results

IMPACT

## **REDUCE THERAPY COST AND IMPROVE OUTCOMES**

identification of new prognostic markers

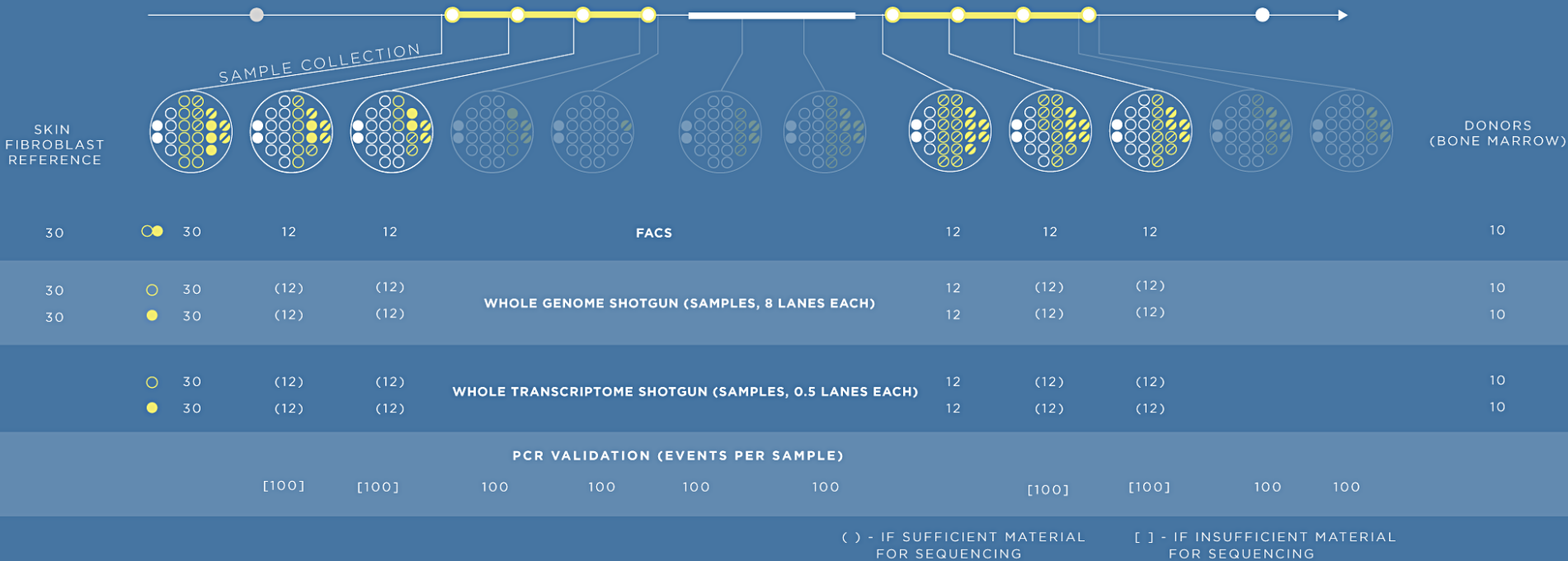
afford earlier detection of high-risk patients to fast-track transplants

discovery of drug targets informed by novel, validated and functionally relevant aberrations

extend to functional characterization and validation of AML mutations in cell lines and animal models

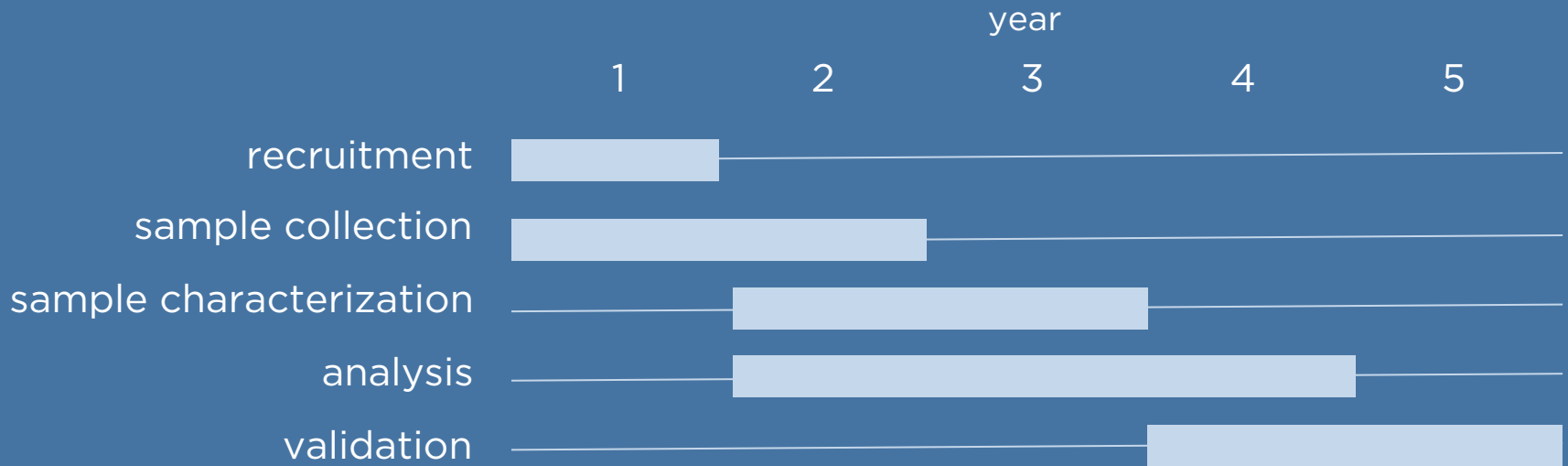
# BUDGET

## SEQUENCING AND VALIDATION SCHEDULE



TIMELINE

**5 YEARS**



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## **PI SUPPORT**

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## **BUDGET AND LOGISTICS**

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